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## INHIBITION OF URIDINE DIPHOSPHOGLUCOSE DEHYDROGENASE BY GALACTOSAMINE-1-PHOSPHATE AND UDP-GALACTOSAMINE

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### SUMMARY

The activity of uridine diphosphoglucose (UDPG) dehydrogenase (UDPG:NAD<sup>+</sup> oxidoreductase, EC 1.1.1.22) from rat liver is inhibited markedly *in vitro* by galactosamine 1-phosphate and UDPgalactosamine. The  $K_i$  value is calculated to be  $6.2 \cdot 10^{-3}$  M for galactosamine-1-phosphate. The type of inhibition is non-competitive. UDPgalactosamine is effective at lower concentrations; a  $K_i$  value of  $6.9 \cdot 10^{-4}$  M has been measured and the inhibition is mainly competitive in nature. This inhibition of UDPG dehydrogenase activity is regarded to be responsible for the decrease of UDPglucuronate *in vivo* after D-galactosamine administration, supporting the significance of galactosamine 1-phosphate and UDPgalactosamine in the metabolic alterations during the development of galactosamine hepatitis.

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### INTRODUCTION

Galactosamine 1-phosphate and UDPgalactosamine accumulate in the liver after the injection of D-galactosamine. The appearance of galactosamine metabolites is the start of a sequence of reactions<sup>1-3</sup> leading to galactosamine hepatitis<sup>4,5</sup> in rats older than three weeks<sup>6</sup>.

In experiments with liver slices, D-galactosamine is a potent inhibitor of glucuronide synthesis<sup>7</sup>. Moreover, it has been shown that D-galactosamine administration causes a marked decrease of UDPglucuronate concentration<sup>8</sup> in rat livers.

The conversion of UDPglucose to UDPglucuronate is catalyzed by uridine diphosphoglucose (UDPG) dehydrogenase (UDPG:NAD<sup>+</sup>oxidoreductase, EC 1.1.1.22). The enzyme is inhibited *in vitro* by UDPxylose<sup>9</sup> and UDPgalactose<sup>10</sup>.

Data presented in this paper show an inhibition of both bovine and rat liver UDPG dehydrogenase by galactosamine 1-P and UDPgalactosamine. This inhibition is regarded as being responsible for the decrease of UDPglucuronate *in vivo* after D-galactosamine administration.

## MATERIAL AND METHODS

UDPG dehydrogenase from male Wistar rats (200–220 g) was purified 80-fold as described by Strominger *et al.*<sup>11</sup>. Calcium phosphate gel adsorption and elution were omitted. For comparison a highly purified UDPG dehydrogenase from bovine liver was obtained from Boehringer GmbH (Mannheim). Enzyme activity was measured spectrophotometrically by following the formation of NADH at 334 nm and 25 °C. Assay mixtures contained, in addition to enzyme, 1.75 mM NAD<sup>+</sup> and 0.01–0.5 mM UDPG in a total volume of 0.85 ml of 0.2 M glycine buffer (pH 8.7)<sup>12</sup>.

The concentrations of galactosamine 1-phosphate and UDPgalactosamine are indicated in the figures.

Galactosamine 1-phosphate was prepared biologically as follows. Male rats received a single injection of 500 mg D-galactosamine-HCl per kg body weight. 1 h after D-galactosamine administration 25% of the injected D-galactosamine had been converted to galactosamine 1-phosphate. The livers were quickly removed under ether anesthesia, transferred to 2.5 vol. chilled 0.9 M HClO<sub>4</sub> and immediately homogenized. The suspensions were centrifuged for 15 min at 18 000 × g, the supernatants being carefully collected, while the sediments were rehomogenized in 1.5 vol. HClO<sub>4</sub>. After a second centrifugation the supernatants were combined and neutralized with KOH. Purification was performed by column<sup>13</sup> and paper chromatography<sup>14</sup>. After hydrolysis, the purity was checked by ion-exchange chromatography<sup>14,15</sup> and paper electrophoresis in 0.1 M borate buffer (pH 8.8). UDPgalactosamine was synthesized enzymatically *in vitro*<sup>14</sup>. The determination of galactosamine metabolites was performed as described previously<sup>14</sup>.

D-Galactosamine-HCl (puriss.) was purchased from C. Roth, OHG (Karlsruhe). All other chemicals were commercial products of analytical grade.

## RESULTS

The Michaelis constants ( $K_m$ ) are calculated to be  $2.0 \cdot 10^{-5}$  M UDPG for rat liver dehydrogenase and  $1.9 \cdot 10^{-5}$  M UDPG for bovine liver dehydrogenase. These

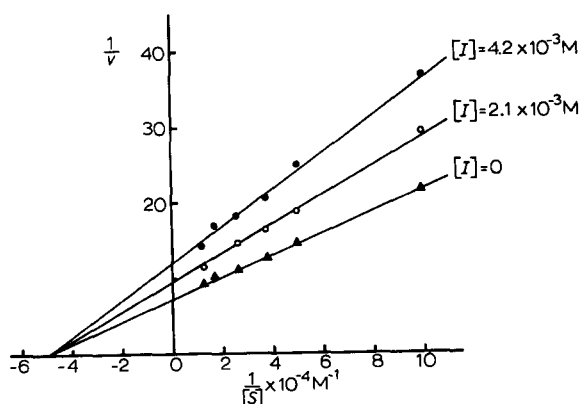


Fig. 1. Lineweaver-Burk plots of rat liver UDPG dehydrogenase activity in the presence of two concentrations of the inhibitor galactosamine 1-phosphate.

values are in agreement with data from the literature<sup>16</sup>. Typical Lineweaver-Burk plots (Fig. 1), obtained with UDPG as substrate and galactosamine 1-phosphate as inhibitor, show that galactosamine 1-phosphate does effectively inhibit the enzyme *in vitro* at concentrations which are present *in vivo* after D-galactosamine administration (see Table I). Measurements with different concentrations of inhibitor have shown that all lines of Lineweaver-Burk transformation cross the ordinate and intersect the negative abscissa. Therefore, the type of inhibition is non-competitive.

TABLE I  
CONCENTRATION OF GALACTOSAMINE METABOLITES

Time-dependent changes of galactosamine 1-phosphate and UDPhexosamine concentrations after intravenous injection of 375 mg D-galactosamine-HCl per kg body weight. The results are expressed as  $\mu\text{moles/g}$  fresh liver,  $\pm\text{S.D.}$  ( $n = 3$ ). The average ratio of UDPgalactosamine to UDPglucosamine is 1:1. Results are expressed in  $\mu\text{moles/g}$ .

| Time after<br>D-galactosamine<br>administration<br>(h) | Sum of<br>galactosamine<br>metabolites | Galactosamine<br>1-phosphate | UDPgalactos-<br>amine | UDPglucos-<br>amine |
|--|--|------------------------------|-----------------------|---------------------|
| 0.5  | 4.4 $\pm$ 0.16                         | 2.9 $\pm$ 0.37               | 0.25 $\pm$ 0.01       | 0.23 $\pm$ 0.01     |
| 1  | 11.5 $\pm$ 0.50                        | 8.5 $\pm$ 0.31               | 0.44 $\pm$ 0.01       | 0.38 $\pm$ 0.01     |
| 3  | 9.9 $\pm$ 0.26                         | 5.6 $\pm$ 0.28               | 0.52 $\pm$ 0.02       | 0.54 $\pm$ 0.01     |
| 6  | 9.4 $\pm$ 0.41                         | 3.8 $\pm$ 0.08               | 0.51 $\pm$ 0.02       | 0.48 $\pm$ 0.02     |
| 12   | 4.5 $\pm$ 0.12                         | 0.4 $\pm$ 0.03               | —                     | —                   |

There is only a slight difference between the  $K_i$  values for the rat ( $6.2 \cdot 10^{-3}$  M) and bovine enzyme ( $7.6 \cdot 10^{-3}$  M) UDPgalactosamine causes an inhibition at concentrations lower than that for the precursor, galactosamine 1-phosphate. At low concentrations of UDPgalactosamine, the inhibition can be overcome by addition of UDPG. The type of inhibition seems to be mainly competitive (Fig. 2). For rat UDPG dehydrogenase a  $K_i$  value of  $6.9 \cdot 10^{-4}$  M was measured.

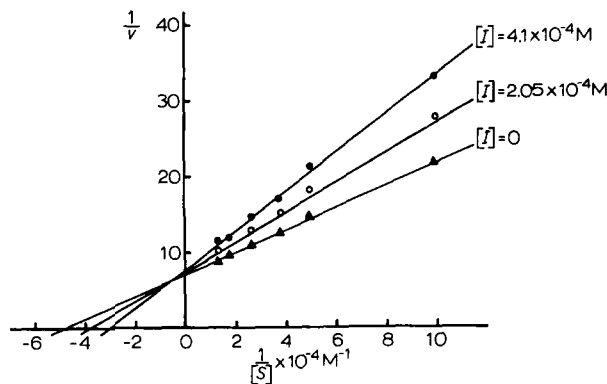


Fig. 2. Double-reciprocal plot of UDPG dehydrogenase activity in the presence of UDPgalactosamine. Each point represents the mean value of duplicate determinations at that concentration.

## DISCUSSION

The enzymatic activity of UDPG dehydrogenase is inhibited by both galactosamine 1-phosphate and UDPgalactosamine *in vitro*. The decreased level of UDPglucuronate in the liver<sup>8</sup> and the inhibition of the synthesis of glucuronides in liver slices<sup>7</sup> can be partially explained by the diminished level of UDPG after D-galactosamine injection<sup>8</sup>. Yet, studies with D-glucosamine instead of D-galactosamine indicate that factors other than the lowered concentration of UDPG must be regarded responsible for the inhibition of UDPglucuronate synthesis<sup>8</sup>. Galactosamine 1-phosphate and UDPgalactosamine reach high levels after D-galactosamine administration and remain at this level for 3 to 6 h. At this time the first signs of a liver injury are seen<sup>3,17</sup>. From the data presented it can be concluded that these metabolites are mainly responsible for the diminished concentration of UDPglucuronate. Inhibition of enzymatic activity by galactosamine metabolites, especially by galactosamine 1-phosphate, has been shown for UDPG pyrophosphorylase (EC 2.7.7.9)<sup>2</sup> supporting the significance of these metabolites in metabolic alterations during the development of galactosamine hepatitis.

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